

# B16F10-Fluc-Neo/iRFP-Puro

## Product Description

Product Name: B16F10-Fluc-Neo/iRFP-Puro  
Catalog Number: CL110  
Lot Number: CL-IM112

Species: Mouse (*Mus musculus*)  
Strain: C57BL/6  
Cell type: Melanoma  
Parental cells: B16F10 (ATCC® CRL-6475™)\*  
Morphology: Epithelial  
Growth mode: Adherent  
Reporter genes: Firefly luciferase (Fluc)  
Near infrared fluorescent protein (iRFP)  
Selection genes: Neomycin (Neo)  
Puromycin (Puro)

This is a polyclonal population derived from the melanoma B16F10 cell line (ATCC® CRL-6475™). Parental B16F10 cells were transduced with 1) LV-Fluc-P2A-Neo (Imanis #LV011) encoding the firefly luciferase (Fluc) cDNA under the spleen focus-forming virus (SFFV) promoter linked to the neomycin resistance gene (Neo) via a P2A cleavage peptide and 2) LV-iRFP-P2A-Puro (Imanis #LV032) encoding the near-infrared fluorescent protein (iRFP; ex/em = 690/713) cDNA under the spleen focus-forming virus (SFFV) promoter linked to the puromycin resistance gene (Puro) via a P2A cleavage peptide. High Fluc and iRFP expressing cells were selected using puromycin. The lentiviral vectors are self-inactivating (SIN) vectors in which the viral enhancer and promoter have been deleted. Transcription inactivation of the LTR in the SIN provirus increases biosafety by preventing mobilization by replication competent viruses and enables regulated expression of the genes from the internal promoters without *cis*-acting effects of the LTR<sup>1</sup>.

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## Mycoplasma Testing

The B16F10-Fluc-Neo/iRFP-Puro cell line has been tested for mycoplasma contamination and is certified mycoplasma free.

## Cell Line Authentication

In light of studies suggesting that 18-36% of cell lines utilized in biomedical research are contaminated or completely misidentified,<sup>2,3</sup> several funding organizations, including NIH, as well as major publishers, including those affiliated with the American Association for Cancer Research (AACR), require cell lines used in research to be authenticated prior to publication<sup>4,5</sup>. The parental B16F10 cell line used to generate B16F10-Fluc-Neo/iRFP-Puro, was authenticated and certified free of interspecies cross contamination by STR profiling with 27 STR loci.

## Recommended Uses

B16F10-Fluc-Neo/iRFP-Puro cells are suitable for *in vitro* and *in vivo* experimentation. B16F10 cells form tumors and pulmonary metastases post implantation into C57BL/6 mice<sup>6</sup>. The Fluc and iRFP transgenes facilitate noninvasive imaging of implanted cells.

## References

- <sup>1</sup>Miyoshi et al. J Virol. 1998. 72:8150-8157.
- <sup>2</sup>Hughes et al. BioTechniques 2007. 43:575-586.
- <sup>3</sup>Chatterjee et al. Science 2007. 315:928-931.
- <sup>4</sup><https://grants.nih.gov/grants/guide/notice-files/NOT-OD-08-017.html>
- <sup>5</sup><http://www.aacrjournals.org/site/InstrAuthors/ifora.xhtml#celllineuse>
- <sup>6</sup>Fidler. Cancer Res, 1975. 35:218-224.

## Storage Instructions

Remove cells from the dry ice packaging and immediately store in the vapor phase above liquid nitrogen (below -130°C).

## Complete Growth Medium

Dulbecco's Modified Eagle's Medium (DMEM)  
10% Fetal bovine serum (FBS)  
1% Penicillin/Streptomycin  
0.8 mg/mL G418 (to maintain high Fluc expression)  
1 µg/mL Puromycin (to maintain high iRFP expression)

## Thawing Instructions

1. Thaw cells by gently swirling in a 37°C water bath. To limit contamination, do not submerge the O-ring and cap.
2. When cells are ~70% thawed (less than 1 min), remove the vial and wipe down with 70% ethanol. Allow tube to dry completely.
3. In a biosafety cabinet, transfer the cells into a 15 mL conical tube containing 5 mL of pre-warmed complete growth medium without selection drugs. Centrifuge cells at ~250 x *g* for 3-5 min.
4. Remove supernatant and resuspend cells in 1 mL complete growth medium without selection drugs. Transfer cells to a T75 flask containing 10 mL pre-warmed complete growth medium without selection drugs.
5. Incubate the culture at 37°C with 5% CO<sub>2</sub>. After 48 hours, replace the culture supernatant with complete growth medium containing 0.8 mg/mL G418 and 1 µg/mL puromycin. Cells should reach full confluency 3-4 days after thawing.

## Subculturing Instructions

Volumes are given for a T75 flask. Increase or decrease as needed. In order to maintain high Fluc and iRFP expression, it is recommended that cells be subcultured in the presence of 0.8 mg/mL G418 and 1 µg/mL puromycin.

1. Remove culture medium from cells.
2. Carefully wash the cell monolayer with 5-10 mL of phosphate buffered saline.
3. Add 2 mL of 0.25% Trypsin-EDTA solution to the flask and incubate at 37°C until cells have dissociated (approx. 2-5 min).
4. Neutralize the trypsin by adding 8 mL complete growth medium, and mix by gently pipetting up and down.
5. Transfer desired portion of the cells to a fresh T75 flask. Add fresh complete growth medium to a total volume of 10 mL and return cells to 37°C/5% CO<sub>2</sub> incubator.

For maintenance, a subcultivation ratio of 1:10 is recommended. At this ratio cells will be ready for passage every 3-4 days. If the growth medium begins to turn brown before cells are ready to be passaged, replace with fresh medium.

## Freezing Medium

Cells can be amplified and used to generate additional frozen stocks. Frozen stocks should be preserved in a designated cryopreservation medium or in complete growth medium without selection drugs supplemented with 5-10% DMSO.

## Additional Considerations

Over trypsinization of B16F10 cells can damage the cells. During trypsinization the cells should be monitored carefully, and the trypsin neutralized immediately upon cell detachment.

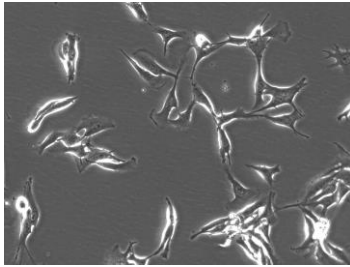
## Certificate of Analysis

Testing performed by Imanis Life Sciences

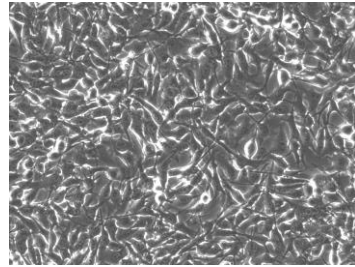
Test description	Result
Post thaw viable cell recovery	Pass QC
Sterility	No contamination detected
Mycoplasma	No contamination detected
Neomycin selection	Pass QC
Puromycin selection	Pass QC
Luciferase expression	Pass QC
Fluorescence expression	Pass QC

## Morphology:

Low density, 200X

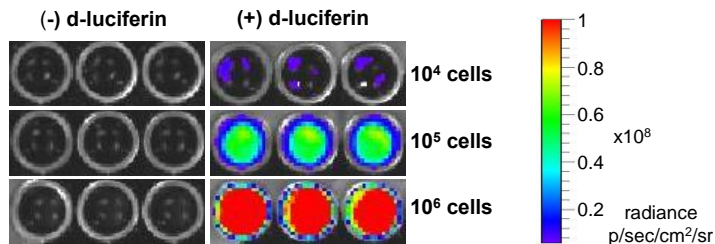


High density, 200X



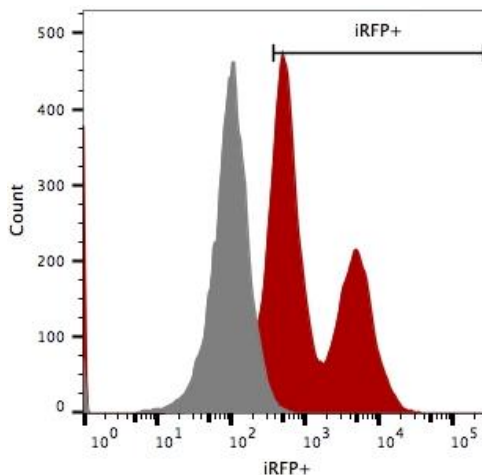
Low and high density photos taken 24 and 48 hours after thawing, respectively.

## Luciferase Expression



$10^4$ ,  $10^5$ , or  $10^6$  cells were placed in wells of a 96-well plate and 3 mg/mL of d-luciferin was added to the indicated wells. The plate was immediately imaged using a Xenogen IVIS Spectrum.

## Fluorescence Expression



B16F10-Fluc-Neo/iRFP-Puro (red) or isotype control (B16F10; grey) cells were fixed with paraformaldehyde and analyzed by flow cytometry.

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Quality Assurance by: SPR

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