

#### **Product Description**

Product Name: Catalog Number: Lot Number:	CL114
Species: Strain:	Mouse ( <i>Mus musculus</i> ) C57BL
Cell type:	Lewis lung carcinoma
Parental cells:	LL/2 (ATCC® CRL-1642 <sup>™</sup> )*
Morphology:	Epithelial

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Growth mode:	Loosely adherent
Reporter gene:	Murine sodium iodide symporter (mNIS)

This is a monoclonal population derived from the Lewis lung carcinoma LL/2 cell line (ATCC® CRL-1642<sup>TM</sup>). Parental LL/2 cells were transduced with LV-mNIS (Imanis #LV008) encoding the murine sodium iodide symporter (mNIS) cDNA under the spleen focus-forming virus (SFFV) promoter. A monoclonal population was isolated following three rounds of selection in a methylcellulose-based semi-solid medium. The lentiviral vector is a self-inactivating (SIN) vector in which the viral enhancer and promoter have been deleted. Transcription inactivation of the LTR in the SIN provirus increases biosafety by preventing mobilization by replication competent viruses and enables regulated expression of the genes from the internal promoters without *cis*-acting effects of the LTR<sup>1</sup>.

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# **Mycoplasma Testing**

The LL/2-mNIS cell line has been tested for mycoplasma contamination and is certified mycoplasma free.

## **Cell Line Authentication**

The parental LL/2 cell line used to generate LL/2-mNIS was authenticated and certified free of interspecies cross contamination by STR profiling with 27 STR loci.

## **Recommended Uses**

LL/2-mNIS cells are suitable for in vitro and in vivo experimentation.

The mNIS protein in not immunogenic in mice. Therefore, these cells can be used for longitudinal imaging studies in both immunocompromised mice and immunocompetent C57BL mice.

#### References

<sup>1</sup>Miyoshi et al. J Virol. 1998. 72:8150-8157.

#### Storage Instructions

Remove cells from the dry ice packaging and immediately store in the vapor phase above liquid nitrogen (below -130°C).

## **Complete Growth Medium**

Dulbecco's Modified Eagle's Medium (DMEM) 10% fetal bovine serum (FBS) 1% Penicillin/Streptomycin

## **Thawing Instructions**

- 1. Thaw cells by gently swirling in a 37°C water bath. To limit contamination, do not submerge the O-ring and cap.
- When cells are ~70% thawed (less than 1 min), remove the vial and wipe down with 70% ethanol. Allow tube to dry completely.
- 3. In a biosafety cabinet, transfer the cells into a 15 mL conical tube containing 5 mL of pre-warmed complete growth medium. Centrifuge at  $\sim$ 250 x g for 3-5 min.
- Remove supernatant and resuspend cells in 1 mL complete growth medium. Transfer cells to a T75 flask containing 10 mL pre-warmed complete growth medium.
- Incubate the culture at 37°C with 5% CO<sub>2</sub>. After 48 hours, replace the culture supernatant with complete growth medium and passage if needed. Cells should reach full confluency 3-4 days after thawing.

## **Subculturing Instructions**

Volumes are given for a T75 flask. Increase or decrease as needed. 1. Remove culture medium from cells.

- 2. Carefully wash the cell monolayer with 5-10 mL of phosphate buffered saline.
- 3. Add 1 mL of 0.25% Trypsin-EDTA solution to the flask and incubate at 37°C until cells have dissociated (approx. 1-2 min).
- 4. Neutralize the trypsin by adding 9 mL complete growth medium and mix by gently pipetting up and down.
- Transfer desired portion of the cells to a fresh T75 flask. Add fresh complete growth medium to a total volume of 10 mL and return cells to 37°C/5% CO<sub>2</sub> incubator.

LL/2 cells are loosely adherent and will begin to detach from flasks prior to reaching 100% confluency. For maintenance, a subcultivation ratio of 1:10 is recommended. At this ratio cells will be ready for passage approximately every 3 days.

## **Freezing Medium**

LL/2-mNIS cells can be amplified and used to generate additional frozen stocks. Frozen stocks should be preserved in a designated cryopreservation medium or in complete growth medium supplemented with 5-10% DMSO.

## **Additional Considerations**

LL/2 cells are loosely adherent. Coating culture plates with poly-D-Lysine prior to use can be used to increase cell adherence to the plates if necessary. Viable cells that have lost adherence can be recovered by low-speed centrifugation and seeded in fresh tissue culture flasks.



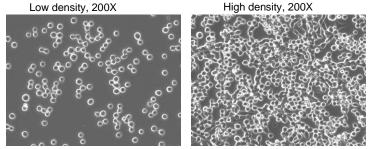
#### **Certificate of Analysis**

Testing performed by Imanis Life Sciences

Test description	Result
Post thaw viable cell recovery	100%
Cells/Vial	~ 5 x 10 <sup>6</sup>
Sterility	No contamination detected
Mycoplasma	No contamination detected
NIS expression	Pass QC
Average Doubling Time	13.7 hours*

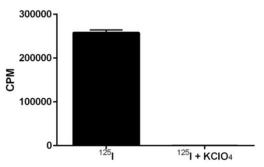
\*Doubling time represents the average doubling time during <u>logarithmic growth</u>. This value should be used for general estimation only.

#### Morphology:



Low and high density photos taken 24 and 72 hours after thawing, respectively.

#### **NIS Expression:**



Uptake of  $^{125}$ l by 3 x 10^5 cells was assayed in the presence or absence of KClO4, an inhibitor of NIS-mediated  $^{125}$ l uptake.

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Quality control by: CDL Quality Assurance by: RLV Effective Date: 15-April-2022