

# **Product Description**

Catalog Number:	CT26.WT-eGFP-Puro CL044 CL-IM26
Species:	Mouse ( <i>Mus musculus</i> )
Strain:	BALB/c
Cell type:	Colorectal carcinoma
Parental cells:	CT26 (ATCC® CRL-2638 <sup>TM</sup> )*
Morphology:	Epithelial
Growth mode:	Adherent
Reporter gene:	Enhanced green fluorescent protein (eGFP)
Selection gene:	Puromycin (Puro)

CT26.WT-eGFP-Puro is a polyclonal population of the murine colorectal cancer CT26.WT cell line transduced with a lentiviral vector encoding the enhanced green fluorescent protein (eGFP) cDNA under the spleen focus-forming virus (SFFV) and the puromycin (Puro) resistance gene under control of the PGK promoter. The lentiviral vector is a self-inactivating (SIN) vector in which the viral enhancer and promoter have been deleted. Transcription inactivation of the LTR in the SIN provirus increases biosafety by preventing mobilization by replication competent viruses and enables regulated expression of the LTR<sup>1</sup>.

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# **Mycoplasma Testing**

This cell line has tested negative for mycoplasma contamination.

# **Cell line Authentication**

Authentication of the parental CT26.WT cell line was confirmed by short tandem repeat (STR) profiling.

#### **Recommended Uses**

eGFP is not recommended for whole animal in-live imaging. Rather, samples can be collected post mortem for analysis by conventional fluorescence microscopy or flow cytometry.

eGFP is immunogenic and may cause tumor rejection in immunocompetent mice. For the most consistent results, immunocompromised mice are recommended for studies.

#### References

<sup>1</sup>Miyoshi et al. J Virol. 1998. 72:8150-8157.

## **Storage Instructions**

Remove cells from the dry ice packaging and immediately store in the vapor phase above liquid nitrogen (below -130°C).

# **Complete Growth Medium**

Dulbecco's Modified Eagle's Medium (DMEM) 10% Fetal bovine serum (FBS) 1% Penicillin/Streptomycin 3 µg/mL Puromycin

Caution! Typical commercial puromycin stocks are provided at a concentration of 10 mg/mL or 10,000X.

Puromycin should <u>NOT</u> be added to the medium until a culture has been well established from the thawed cells (about 1 week). It is also recommended that a backup frozen cell stock be generated (see below) before adding puromycin to the growth medium.

## **Thawing Instructions**

- 1. Thaw cells by gently swirling in a 37°C water bath. To limit contamination, do not submerge the O-ring and cap.
- 2. When cells are ~70% thawed (~1 min), remove the vial and wipe down with 70% ethanol. Allow tube to dry completely.
- 3. In a biosafety cabinet, transfer the cells into a 15 mL conical tube containing 5 mL of pre-warmed complete growth medium. Centrifuge cells at ~250 x g for 3-5 min.
- Remove supernatant and resuspend cells in 1 mL complete growth medium. Transfer cells to a T75 flask containing 10 mL pre-warmed complete growth medium.
- Incubate the culture at 37°C with 5% CO<sub>2</sub>. Cells should reach full confluency 1-2 days after thawing.

## Subculturing Instructions

Volumes are given for a T75 flask. Increase or decrease as needed. 1. Remove culture medium from cells.

- 2. Carefully wash the cell monolayer with 5-10 mL of phosphate buffered saline.
- 3. Add 2 mL of 0.25% Trypsin-EDTA solution to the flask and incubate at room temperature until cells have dissociated.
- 4. Neutralize the trypsin by adding 8 mL complete growth medium, and mix by gently pipetting up and down.
- 5. Transfer desired portion of the cells to a fresh T75 flask. Add fresh complete growth medium to a total volume of 10 mL and return cells to 37°C/5% CO<sub>2</sub> incubator.

For maintenance, a subcultivation ratio of 1:10 is recommended. At this ratio cells will be ready for passage approximately every 3-4 days.

## **Freezing Medium**

Cells can be amplified and used to generate additional frozen stocks. Frozen stocks should be preserved in a designated cryopreservation medium or in complete growth medium <u>without puromycin</u> supplemented with 5-10% DMSO.

# **Additional Considerations**

Cells easily detach from tissue culture surfaces. Coating culture pates with poly-D-Lysine prior to use is recommended for applications involving frequent washing or rocking. These cells will not form 100% confluent monolayers; the cells are ready for passage when they reach 80-90% confluency.

# CT26.WT-eGFP-Puro

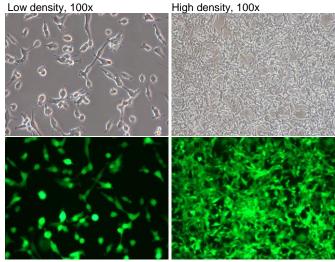


## **Certificate of Analysis**

#### Testing performed by Imanis Life Sciences:

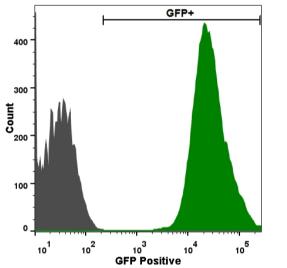
Test description	Result
Post thaw viable cell recovery	Pass QC
Sterility	No contamination detected
Mycoplasma	No contamination detected
Puromycin selection	Pass QC
Fluorescence expression	Pass QC

#### Morphology



Low and high density photos taken at various times after thawing.

#### Fluorescence Expression:



CT26.WT-eGFP-Puro (green) or isotype control (CT26.WT-mNIS; grey) cells were fixed with paraformaldehyde and analyzed by flow cytometry.

#### Legal Disclaimers

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