

Product Description

Product Name: Catalog Number: Lot Number:	LL/2-eGFP-Puro CL051 CL-IM35
Species:	Mouse (<i>Mus musculus</i>)
Strain:	C57BL
Cell type:	Lewis lung carcinoma
Parental cells:	LL/2 (ATCC [®] CRL-1642 [™])*
Morphology:	Epithelial
Growth mode:	Loosely adherent
Reporter gene:	Enhanced green fluorescent protein (eGFP)
Selection gene:	Puromycin (Puro)

This is a polyclonal population derived from the Lewis lung carcinoma LL/2 cell line (ATCC® CRL-1642TM). Parental LL/2 cells were transduced with a LV-eGFP-PGK-Puro (Imanis #LV031) encoding the enhanced green fluorescent protein (eGFP) cDNA under the spleen focus-forming virus (SFFV) promoter and the puromycin resistance gene (Puro) under the phosphoglycerate kinase (PGK) promoter. High eGFP expressing cells were selected using puromycin. The lentiviral vectors are self-inactivating (SIN) vectors in which the viral enhancer and promoter have been deleted. Transcription inactivation of the LTR in the SIN provirus increases biosafety by preventing mobilization by replication competent viruses and enables regulated expression of the genes from the internal promoters without *cis*-acting effects of the LTR¹. * The ATCC trademark and any and all ATCC catalog numbers are trademarks of the American Type Culture Collection

Mycoplasma Testing

This cell line has tested negative for mycoplasma contamination.

Cell line Authentication

Authentication of the parental LL/2 cell line was confirmed by short tandem repeat (STR) profiling.

Recommended Uses

These cells are suitable for in vitro and in vivo experimentation.

eGFP is not recommended for whole animal in-live imaging. Rather, samples can be collected post mortem for analysis by conventional fluorescence microscopy or flow cytometry.

eGFP is immunogenic and may cause tumor rejection in immunocompetent mice. For the most consistent results, immunocompromised mice are recommended for studies.

References

¹Miyoshi et al. J Virol. 1998. 72:8150-8157.

Storage Instructions

Remove cells from the dry ice packaging and immediately store in the vapor phase above liquid nitrogen (below -130°C).

Complete Growth Medium

Dulbecco's Modified Eagle's Medium (DMEM) 10% fetal bovine serum (FBS) 1% Penicillin/Streptomycin 2 µg/mL puromycin

Caution! Typical commercial puromycin stocks are provided at a concentration of 10 mg/mL or 10,000X.

Puromycin should <u>NOT</u> be added to the medium until a culture has been well established from the thawed cells (about 1 week). It is also recommended that a backup frozen cell stock be generated (see below) before adding puromycin to the growth medium.

Thawing Instructions

- 1. Thaw cells by gently swirling in a 37°C water bath. To limit contamination, do not submerge the O-ring and cap.
- 2. When cells are ~70% thawed (~1 min), remove the vial and wipe down with 70% ethanol. Allow tube to dry completely.
- 3. In a biosafety cabinet, transfer the cells into a 15 mL conical tube containing 5 mL of pre-warmed complete growth medium. Centrifuge cells at ~250 x g for 3-5 min.
- Remove supernatant and resuspend cells in 1 mL complete growth medium. Transfer cells to a T75 flask containing 10 mL pre-warmed complete growth medium.
- Incubate the culture at 37°C with 5% CO₂. Cells should reach full confluency 1-2 days after thawing.

Subculturing Instructions

Volumes are given for a T75 flask. Increase or decrease as needed.

- 1. Remove culture medium from cells.
- 2. Carefully wash the cell monolayer with 5-10 mL of phosphate buffered saline.
- Add 2 mL of 0.25% Trypsin-EDTA solution to the flask and incubate at room temperature until cells have dissociated (approx. 2-5 min).
- 4. Neutralize the trypsin by adding 8 mL complete growth medium, and mix by gently pipetting up and down.
- Transfer desired portion of the cells to a fresh T75 flask. Add fresh complete growth medium to a total volume of 10 mL and return cells to 37°C/5% CO₂ incubator.

These cells should be passaged when they reach ~80% confluency. Beyond this density, the cells begin to lift off the plate or clump together in large balls. Some of these cells remain viable and can be collected by centrifugation and re-plated if needed. For maintenance, a subcultivation ratio of 1:10 is recommended. At this ratio cells will be ready for passage every 3-4 days.

Freezing Medium

LL/2-eGFP-Puro cells can be amplified and used to generate additional frozen stocks. Frozen stocks should be preserved in a designated cryopreservation medium or in complete growth medium <u>without puromycin</u> supplemented with 5-10% DMSO.

Additional Considerations

LL/2 cells are loosely adherent. Coating culture plates with poly-D-Lysine prior to use can be used to increase cell adherence to the plates if necessary.



Certificate of Analysis

Testing performed by Imanis Life Sciences

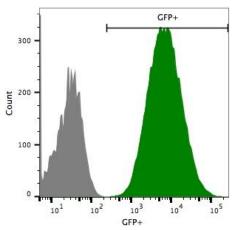
Test description	Result
Post thaw viable cell recovery	Pass QC
Sterility	No contamination detected
Mycoplasma	No contamination detected
Puromycin selection	Pass QC
Fluorescence expression	Pass QC

Morphology:

High density, 200X Low density, 200X ЬÖ 6

Low and high density photos taken at various times after thaw.

Fluorescence Expression



LL/2-eGFP-Puro (green) or isotype control (LL/2-Fluc-Puro; grey) cells were fixed with paraformaldehyde and analyzed by flow cytometry.

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