

## Product Description

Product Name: B16F10-Fluc-hNIS  
Catalog Number: CL139  
Lot Number: CL-IM153

Species: Mouse (*Mus musculus*)  
Strain: C57BL/6  
Cell type: Melanoma  
Parental cells: B16F10 (ATCC® CRL-6475™)\*  
Morphology: Epithelial  
Growth mode: Adherent  
Reporter genes: Firefly luciferase (Fluc)  
Human sodium iodide symporter (hNIS)

This is a population derived from the melanoma B16F10 cell line (ATCC® CRL-6475™). Parental B16F10 cells were transduced with LV-Luc2-P2A-hNIS (Imanis #LV023) encoding the firefly luciferase (Luc2; Fluc) cDNA under the spleen focus-forming virus (SFFV) promoter and linked to the human sodium iodide symporter (hNIS) cDNA via a P2A cleavage peptide. High Fluc and hNIS expressing cells were selected using a methylcellulose-based semi solid medium. The lentiviral vector is a self-inactivating (SIN) vector in which the viral enhancer and promoter have been deleted. Transcription inactivation of the LTR in the SIN provirus increases biosafety by preventing mobilization by replication competent viruses and enables regulated expression of the genes from the internal promoters without *cis*-acting effects of the LTR<sup>1</sup>.

\* The ATCC trademark and any and all ATCC catalog numbers are trademarks of the American Type Culture Collection

## Mycoplasma Testing

This cell line has been tested for mycoplasma contamination and is certified mycoplasma free.

## Cell Line Authentication

The parental B16F10 cell line was authenticated and certified free of interspecies cross contamination by STR profiling.

## Recommended Uses

These cells are suitable for *in vitro* and *in vivo* experimentation.

The Fluc transgene facilitates *in vivo* noninvasive bioluminescent imaging of implanted cells. The hNIS transgene facilitates high resolution, 3D SPECT/PET imaging of implanted cells. Fluc is immunogenic and may cause tumor rejection in immunocompetent mice. For the most consistent results, immunocompromised mice are recommended for studies.

## References

<sup>1</sup>Miyoshi et al. J Virol. 1998. 72:8150-8157.

## Storage Instructions

Remove cells from the dry ice packaging and immediately store in the vapor phase above liquid nitrogen (below -130°C).

## Complete Growth Medium

Dulbecco's Modified Eagle's Medium (DMEM)  
10% fetal bovine serum (FBS)  
1% Penicillin/Streptomycin

## Thawing Instructions

1. Thaw cells by gently swirling in a 37°C water bath. To limit contamination, do not submerge the O-ring and cap.
2. When cells are ~70% thawed (less than 1 min), remove the vial and wipe down with 70% ethanol. Allow tube to dry completely.
3. In a biosafety cabinet, transfer the cells into a 15 mL conical tube containing 5 mL of complete growth medium. Centrifuge cells at ~250 x g for 3-5 min.
4. Remove supernatant and resuspend cells in 1 mL complete growth medium. Transfer cells to a T75 flask containing 10 mL complete growth medium.
5. Incubate the culture at 37°C with 5% CO<sub>2</sub>. After 48 hours, replace the culture supernatant with fresh complete growth medium. Cells should reach full confluency 3-4 days after thawing.

## Subculturing Instructions

Volumes are given for a T75 flask; increase or decrease as needed.

1. Remove culture medium from cells.
2. Carefully wash the cell monolayer with 5-10 mL of phosphate buffered saline.
3. Add 2 mL of 0.25% Trypsin-EDTA solution to the flask and incubate at 37°C until cells have dissociated (approx. 2-3 min).
4. Neutralize the trypsin by adding 8 mL complete growth medium, and mix by gently pipetting up and down.
5. Transfer desired portion of the cells to a fresh T75 flask. Add fresh complete growth medium to a total volume of 10 mL and return cells to 37°C/5% CO<sub>2</sub> incubator.

For maintenance, a subcultivation ratio of 1:10 is recommended. At this ratio cells will be ready for passage every 2-4 days.

## Freezing Medium

These cells can be amplified and used to generate additional frozen stocks. Cryopreservation of low passage stocks is recommended. Frozen stocks should be preserved in a designated cryopreservation medium or in complete growth medium without antibiotics supplemented with 5-10% DMSO.

## Additional Considerations

B16F10 cells produce melanin; accumulation of melanin turns the cells and culture medium dark brown or black. Melanin is toxic and B16F10 cells will die in the presence of excess melanin. Culture medium should be changed as soon as it becomes black, even if the cells are not confluent. Typically, media changes between passages are not required.

## Certificate of Analysis

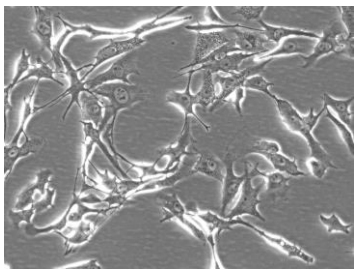
Testing performed by Imanis Life Sciences

Test description	Result
Post thaw viable cell recovery	98%
Cells per vial	~ 5.5 x 10 <sup>6</sup>
Sterility	No contamination detected
Mycoplasma	No contamination detected
Luciferase expression	Pass QC
NIS expression	Pass QC
Average Doubling Time	21.1 hours*

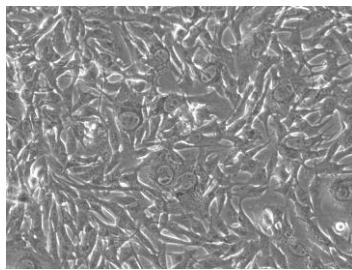
\*Doubling time represents the average doubling time during logarithmic growth. This value should be used for general estimation only.

## Morphology

Low density, 200X

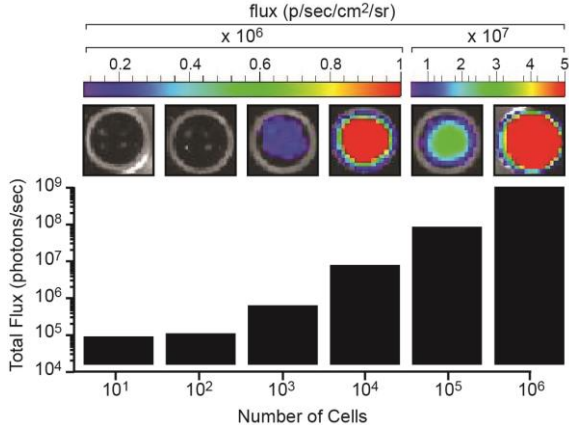


High density, 200X



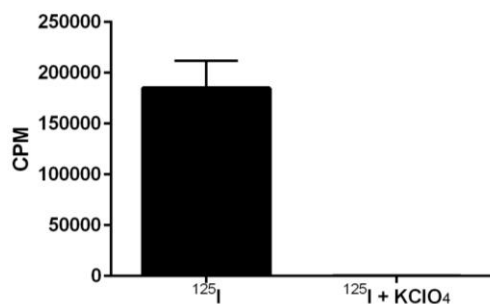
Low and high density photos taken at various times after thawing.

## Luciferase Expression



The indicated number of cells were placed in wells of a 96-well plate. After the addition of 3 mg/mL d-luciferin, the plate was immediately imaged using an IVIS Spectrum. The total flux (photons/sec) was plotted as a function of cell number.

## NIS Expression



Uptake of <sup>125</sup>I by 3 x 10<sup>5</sup> cells was assayed in the presence or absence of KClO<sub>4</sub>, an inhibitor of NIS-mediated <sup>125</sup>I uptake.

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Effective Date: 26-Oct-2022