# LV-Luc2-P2A-EmGFP



## **Product Description**

Product Name: LV-Luc2-P2A-EmGFP Catalog Number: LV050-L (1 mL)

Lot Number: IML012

Reporter genes: Firefly luciferase (Luc2)

Emerald green fluorescent protein (EmGFP)

Promoter: Spleen focus-forming virus (SFFV)

Quantity: 1 mL (L)

Titer: 2.35 x 10<sup>7</sup> TU/mL\* Storage media: Serum-free media

Shipping: Dry ice

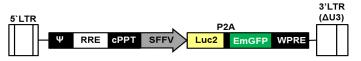
Storage: Store at ≤ -70°C upon receipt. Freeze-thaw

cycles will decrease titer.

Shelf life: One year from date of receipt under proper

storage conditions.

This is a ready-to-use lentivirus preparation. The virus encodes the firefly luciferase (Luc2) cDNA linked to the emerald green fluorescent protein (EmGFP) cDNA via a P2A cleavage peptide; the spleen focus-forming virus (SFFV) promoter transcriptionally controls expression of the Luc2-P2A-EmGFP cDNA (see below). The lentiviral vectors are self-inactivating (SIN) vectors in which the viral enhancer and promoter have been deleted. Transcription inactivation of the LTR in the SIN provirus increases biosafety by preventing mobilization by replication competent viruses and enables regulated expression of the genes from the internal promoters without *cis*-acting effects of the LTR<sup>1</sup>.



5' LTR: 5' long terminal repeat ψ: RNA packaging signal RRE: Rev response element cPPT: Central polypurine tract

SFFV: Spleen focus-forming virus promoter

Luc2: Firefly luciferase P2A: P2A cleavage peptide

EmGFP: Emerald green fluorescent protein

WPRE: Woodchuck hepatitis virus posttranscriptional regulatory element

3' LTR/ΔU3: 3' self-inactivating long terminal repeat

### \*Titration by dPCR:

A PSI  $(\psi)$  probe-based dPCR assay was used to measure the number of copies of lentiviruses stably integrated into the genome after transduction of HeLa H1 cells (transducing units per mL).

Traditional p24 ELISA titrations measure both functional and non-functional lentivirus particles. However, this method overestimates the functional titer, as the p24 protein pool includes a variable amount of free p24 and p24 associated with non-functional vector particles. This ratio can vary greatly between each lot, so the titration is inherently inaccurate. While qPCR titers may appear lower than p24 ELISA, they are more accurate and functional.

#### **Safety Precaution:**

All culture work with lentiviruses should be performed by trained personnel and performed under BSL2 containment following NIH guidelines.



#### **Basic Lentivirus Transduction Protocol**

Volumes are given for a 6-well plate; increase or decrease as needed. See the Transduction Tips section for additional considerations/modifications.

- 1. Seed cells in complete medium at an appropriate density to achieve 60-70% confluency the next day (e.g.  $\sim 2.5 \times 10^5$  HeLaH1 cells). Incubate cells overnight in a 37°C/5% CO<sub>2</sub> incubator.
- 2. Thaw lentivirus stock on ice.
- In a microcentrifuge tube, dilute lentivirus to 1 mL total in serum free media. (See tips below for notes about determining optimal MOIs.)
- 4. Remove culture medium from cells and replace with prepared lentivirus.
- 5. Return cells to 37°C/5% CO2 incubator.
- 6. After 4 hours add 1 mL complete medium to each well and return cells to 37°C/5% CO<sub>2</sub> incubator.
- 7. 3 days after transduction, check transgene expression according to an appropriate protocol.

## **Transduction Tips**

- To determine the optimal MOI for transductions: plate several wells of the target cells and infect with increasing MOIs (e.g 1, 3, 10, and 30). Typically, primary cells require higher MOIs than established cell lines.
- 2. Polybrene® (Imanis #REA001) can be added to the transduction mixture to enhance transduction efficiency². The final concentration of Polybrene® in the transduction medium should be 4-8 μg/mL. Polybrene® can be cytotoxic to some cells and it is not advisable to incubate these cells with Polybrene® overnight; the transduction cocktail may be removed after 3-4 h and replaced with complete media.
- 3. A spin infection can also be used to increase transduction efficiency<sup>3</sup>. Once the transduction mixture is added to the cells, centrifuge the plate at 800 x g for 30 min at room temperature, before placing the cells in a 37°C/5%CO<sub>2</sub> incubator.
- 4. The presence of serum in the transduction mixture can greatly affect transduction efficiency<sup>4</sup>. In general, lower serum concentrations result in higher transduction efficiencies, though optimizing serum concentrations is recommended for each cell type.

### References

<sup>1</sup>Miyoshi et al. J Virol. 1998. 72:8150-8157.

<sup>2</sup>Konopka et al. J Gen Virol 1991. 72: 2685-2696.

<sup>3</sup>O'Doherty et al. J Virol. 2000. 74:10074-10080.

<sup>4</sup>Andreadis and Palsson. Human Gene Ther. 1997. 8:285-291.

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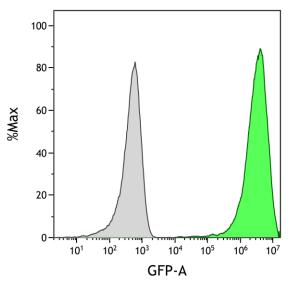


### **Certificate of Analysis**

Testing performed by Imanis Life Sciences

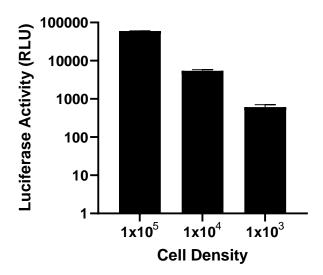
Test description	Result
Virus titer	2.35 x 10 <sup>7</sup> TU/mL
Sterility	No contamination detected
Luciferase expression	Pass QC
Fluorescence expression	Pass QC

#### Fluorescence Expression:



HEK-293T cells were transduced with LV-Luc2-P2A-EmGFP (MOI=10). The transduced cells (green) and untransduced control cells (grey) were analyzed by flow cytometry.

## **Luciferase Expression:**



HEK-293T cells were transduced with LV-Luc2-P2A-EmGFP at an MOI of 10. To measure luciferase expression, 10<sup>5</sup>, 10<sup>4</sup>, or 10<sup>3</sup> cells were seeded in wells of a 96-well plate and 15 mg/mL of d-luciferin was added. Luciferase activity was immediately measured using a TECAN plate reader.

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For in vitro & In vivo use. This certificate is a declaration of analysis at the time of manufacture.

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